

## Annex A

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## Annex B

### Agenda

1. Introduction (including arrangements for Workshop)
2. Adoption of Agenda
3. Keynote presentations
  - 3.1 Individual variation in contaminant levels
  - 3.2 Xenobiotics and metabolism
  - 3.3 Cancer in cetaceans, a potential biomarker of environmental contamination
  - 3.4 Epidemiology/epizootics and contaminants
  - 3.5 Significance and potential of biomarkers in marine mammal toxicology
4. Effects of chemical pollution on cetaceans
  - 4.1 Direct effects
    - 4.1.1 Lethal effects
    - 4.1.2 Sub-lethal effects
      - 4.1.2.1 Diseases
      - 4.1.2.2 Reproduction and early development
      - 4.1.2.3 Immune suppression
      - 4.1.2.4 Cancer induction and mutagenic effects
      - 4.1.2.5 Behaviour
      - 4.1.2.6 Epizootics
  - 4.2 Indirect effects
5. Research implications
  - 5.1 Consideration of synergistic/cumulative effects
  - 5.2 Exotic compounds
  - 5.3 Adequacy of present monitoring
    - 5.3.1 Sources of samples
    - 5.3.2 Biomarkers
    - 5.3.3 Biological variables
    - 5.3.4 Pathology examinations
    - 5.3.5 Specimen banking
  - 5.4 Further evaluation of the relationship between toxic burden and impacts
  - 5.5 Risk and hazard assessment techniques
  - 5.6 Trends in global contamination
  - 5.7 Identification of additional areas of concern
  - 5.8 Review of submitted proposals
6. Implications for the future work of the IWC Scientific Committee
7. Recommendations
8. Publications
9. Adoption of Report

## Annex C

### List of Documents

#### SC/M95

SC/M95/P1. BOON, J.P. Monitoring concentrations of organochlorines, and biochemical and immunological effects in samples of living marine mammals. An introductory note on the possibilities.

SC/M95/P2. MOORE, M.J., MILLER, C.A., WHITE, R.D., SHEA, D., WEISBROD, A.V. and STEGEMAN, J.J. Histological and cytochrome P4501A expression in tissues of pilot whales, *Globicephala melaena*, stranded on Cape Cod, MA USA.

SC/M95/P3. CAURANT, F. Cadmium and mercury in pilot whales: physico-chemical forms of storage and potential hazard to the species.

SC/M95/P4. COLBORN, T. and SMOLEN, M. An epidemiological analysis of persistent organochlorine contaminants in large cetaceans.

SC/M95/P5. CRAIG, A.M., ORPIN, C.G. and BLYTHE, L.L. Biotransformation of marine pollutants, particularly crude oil alkanes, by forestomach bacteria from the bowhead whale. [Research Proposal.]

SC/M95/P6. AGUILAR, A., BORRELL, A. and PASTOR, T. Factors affecting variability of persistent pollutant levels in cetaceans.

SC/M95/P7. BORRELL, A. Summary of temporal trends in pollutant levels observed in marine mammals.

SC/M95/P8. BROUWER, A. Metabolism of xenobiotics in laboratory animals and wildlife species: potential impact on physiology and health.

SC/M95/P9. PEAKALL, D.B. Biomarkers as pollution indicators with special reference to cetaceans.

SC/M95/P10. MARTINEAU, D., LAIR, S., DE GUISE, S. and BELAND, P. Cancer in cetaceans, a potential biomarker for environmental contamination.

SC/M95/P11. SIMMONDS, M.P. Marine mammal mass mortality events: Environmental influences and science.

SC/M95/P12. DONOVAN, G.P. Pollution references on the IWC database.

SC/M95/P13. TANABE, S., AONO, S., FUJISE, Y., KATO, H. and TATSUKAWA, R. Persistent organochlorine residues in the Antarctic minke whale, *Balaenoptera acutorostrata*.

SC/M95/P14. FISHERIES AGENCY GOVERNMENT OF JAPAN. Project of global environment monitoring with fishing vessel network.

SC/M95/P15. KENNEDY, S. Morbillivirus epizootics in aquatic mammals.

**SC/46**

SC/46/O 8. REIJNDERS, P.J.H. Contaminants and cetaceans: Reasons for concern?

SC/46/O 12. JONES, P.D., HANNAH, D.J., BUCKLAND, S.J., VAN MAANEN, T., LEATHEM, S.V., DAWSON, S., SLOOTEN, E., VAN HELDEN, A. and DONOGHUE, M. Planar chlorinated hydrocarbons in New Zealand marine mammals.

SC/46/O 14. MOSCROP, A. and SIMMONDS, M.P. The significance of pollution for marine cetaceans.

SC/46/O 16. BROWN, K. Requirements for a comprehensive assessment of pollution in cetaceans: quantification, evaluation and absolute threat of pollutants.

SC/46/O 20 (revised). BOWLES, D. An overview of the concentrations and effects of heavy metals in cetacean species.

SC/46/RP4. CAURANT, F., BLOCH, D., MOREAU, A., BALLAN-DUFRANCAIS, C. and ALGOET, M. Histo-pathology of kidney and liver tissues of the pilot whales off the Faroe Islands, related with high levels of cadmium and mercury. [Research Proposal.]



## Annex D

### Glossary

**Aromatic hydrocarbon (Ah) receptor.** A protein that binds dioxins, dibenzofurans, non-*ortho* and mono-*ortho* PCBs, and 4 and 5 ring hydrocarbons.

**Assays.** Procedure for measurement or identification.

**Biomarker.** A biological response to a chemical or chemicals that give a measure of exposure and sometimes, also of toxic effect.

**PCB Congeners.** One of the 201 chemical forms of the PolyChlorinated Biphenyls. These are identified by numbers, e.g. OH-PCB-77, hydroxylated polychlorinated biphenyl, number 77.

**Cytochrome P450 1A (CYP-1A).** An enzyme found in epithelia and endothelia, that is induced by and metabolises dioxin, dibenzofurans, some PCBs, and 4 and 5 ring hydrocarbons. The protein concentration is commonly measured as a biomarker for the exposure and effect of those compounds.

**DNA adducts.** Covalent binding of pollutants, especially the polynuclear aromatic hydrocarbons (PAHs), to DNA. This clearly indicates exposure of the organism to the pollutant.

**Endocrine disruptor.** Any compound that interacts with reproductive physiology to alter normal expression of sexually dimorphic form and/or function.

**Ethoxyresorufin-O-deethylase (EROD).** A model substrate for metabolism by CYP-1A. The EROD activity is used in conjunction with CYP-1A as a biomarker of exposure to and effect of Ah receptor activating compounds.

**Halogenated aromatic hydrocarbons (HAHs).** Synthetic compounds containing chlorine, fluorine or bromine atoms, such as PCBs, pesticides and brominated fire retardants.

**Hazard assessment.** Estimation of potential risk.

**Hepa1c1c7.** Liver tumour cell line.

**Immunosuppression.** A measureable alteration in any component(s) of the immune system that is likely to result in increased susceptibility to disease.

**Mixed function oxidases (MFOs).** A group of enzymes that are capable of metabolising a wide range of natural and unnatural chemicals. The function of these enzymes is to increase excretion of potentially harmful chemicals, but in the case of man-made compounds the metabolites may be more toxic than the original compound.

**OH-PCB-77.** Hydroxylated polychlorinated biphenyl, number 77.

**Persistent induction of biotransformation enzymes.** Continuous triggering of metabolism enzymes.

**Polychlorinated biphenyls (PCBs).** Synthetic compounds used for electrical capacitors in the 1950s to 1970s.

**Polycyclic aromatic hydrocarbons (PAHs).** Large groups of naturally occurring aromatic compounds containing two or more benzene rings fused together. Also, oil compounds derived directly (petrogenic) or indirectly via combustion (pyrogenic). Some of these compounds, such as benzo(*a*)pyrene, are known carcinogens.

**Risk assessment.** Estimation of extent of risk.

**Residue levels.** Concentrations of contaminants in substrate (e.g. soil, fluid, tissue).

**T<sub>4</sub>-binding competition on TTR.** Hepatic thyroxine binding competition on transthyretin.

**Xenobiotics.** Foreign substances to a living organism (e.g. contaminants).

## ABBREVIATIONS OF CHEMICAL COMPOUNDS

3,3',4,4'-TeCB - tetrachlorobiphenyl.

CPs - chlorinated paraffins.

HCB - hexachlorobenzene.

HCHs - hexachlorocyclohexanes.

MSF - methylsulphate.

MSF-PCBs - methylsulphate-PCBs.

OC - organochlorines.

OH - organohalogens.

OH-PCBs - hydroxy-PCBs.

PAH - polycyclic aromatic hydrocarbons.

PBBs - polybrominated biphenyls.

PBDEs - polybrominated diphenylethers.

PCBs - polychlorinated biphenyls.

PCDDs - polychlorinated dibenzo-*p*-dioxins.

PCDEs - polychlorinated diphenylethers.

PCDFs - polychlorinated dibenzofurans.

PCNs - polychlorinated naphthalenes.

PCTs - polychlorinated terphenyls.

TBT - tributyl tin.

TCP - tris (*G*-chlorophenyl) methanol.

TCPMe - tris (*G*-chlorophenyl) methane.

TPT - triphenyl tin.

## Annex E

### Priorities in Pathology

D. Martineau

#### INTRODUCTION

The following protocol is intended to further research into the chronic toxicity of organochlorinated compounds (OC) in cetaceans. OC toxicity has been well documented in domestic and laboratory animals. These compounds are known to induce squamous metaplasia of various glands (including mammary glands), to be oestrogenic and to produce mucous metaplasia in glandular stomachs and gastric erosions. A general record of each animal should be kept, including age, sex, reproductive status and, when available, the cause of death. Relevant OC contamination levels in the examined animal should be obtained. The severity and nature of the lesions might be ultimately correlated with OC levels in order to obtain a 'dose-response' relation.

It must be stressed that this protocol is not designed to determine the cause of death. Whenever possible, one should seek ideal conditions for post-mortem examination. These should include a complete, careful examination performed by trained personnel in an appropriate facility. In many field situations, these requirements cannot be fulfilled. Further information is provided in Kuiken and García Hartmann (1993).

#### GENERAL PROCEDURE

Thin, flat sections (approximately 3mm thick, 1cm wide and 2cm long) should be cut and placed in neutral buffered 10% formalin.

Abnormalities: the size, shape, number and colour of any abnormalities should be described using simple terms, for example: 'a dozen randomly distributed, (1cm-diameter) round, dark red areas surrounded by a pale (2mm thick) halo are present on the lower aspect of the heart'.

#### SAMPLES REQUIRED FOR DETECTION OF CHRONIC OC TOXICITY

*Thyroid glands:* cut gland parallel to its long axis, before fixation.

*Lungs:* total of six sections:

Left lung:	cranial	Right lung:	cranial
	middle		middle
	caudal		caudal

*Stomach:* a section of the mucosa of each stomach. If ulcers are present, a section of the lesion and a section of normal bordering tissue.

*Liver:* three sections: middle and both sides.

*Kidney:* three sections of each: cranial, middle, caudal.

*Adrenal glands:* cut both glands parallel to their long axis, before fixation.

*Mammary glands*: three sections of each: cranial, middle, caudal.

*Uterus*: three transverse sections of each uterine horn at various levels (total of six sections).

#### REFERENCE

Kuiken, T. and García Hartmann, M. 1993. Proceedings of the First European Cetacean Society Workshop on Cetacean Pathology: dissection techniques and tissue sampling. *ECS Newsletter* 17:1-39.

