Diversity, habitat associations and stock structure of odontocete cetaceans in the waters of American Samoa, 2003-06

DAVID W. JOHNSTON+*, JOOKE ROBBINS#, MARIE E. CHAPLA*, DAVID K. MATTILA~ AND KIMBERLY R. ANDREWS++

ABSTRACT

Little is known about the species composition, distribution, abundance or stock structure of odontocetes in the central and western tropical Pacific Ocean, including those inhabiting the US Exclusive Economic Zone (EEZ) waters of American Samoa. While some information on species presence in this region has been gleaned from anecdotal sightings and whaling and stranding records, odontocete diversity in the waters of American Samoa has never been formally investigated. This lack of information precludes efforts to determine the sustainability of cetacean populations within US EEZ waters. This paper reports on the first dedicated surveys to document the presence and distribution of odontocete cetaceans in the waters of American Samoa. A series of small-boat photo-identification and biopsy surveys for cetaceans were conducted in the nearshore waters of Tutuila during 2003-06. In addition, ship-based visual surveys were conducted in the waters surrounding the Manu'a Islands, Rose Atoll and Swains Island in summer 2006. A total of 58 groups of odontocete cetaceans were encountered during both small-boat and ship-based surveys: spinner dolphins (Stenella longirostris, n=34), rough-toothed dolphins (Stene bredanensis, n=10, sperm whales (Physeter macrocephalus, n=3), false killer whales (Pseudorca crassidens, n=5), bottlenose dolphins (Tursiops truncatus., n=1), dwarf sperm whales (Kogia sima, n=1), short-finned pilot whales (Globicephala macrorhynchus, n=1), and three groups of unidentified odontocetes. Photographs were analysed for quality and individuals with distinctive markings were selected for entry into a photo-identification catalogue. The resultant catalogue included 46 spinner dolphins, 41 rough-toothed dolphins, 2 bottlenose dolphins, 5 false killer whales, 4 pilot whales, 1 dwarf sperm whale and 4 sperm whales. Thirteen spinner dolphins and 14 rough-toothed dolphins were sighted in multiple years. To investigate stock structure, spinner dolphin genetic data were used to compare mitochondrial control region genetic diversity and allele frequencies between American Samoa and the Hawaiian Islands. American Samoa had a higher genetic diversity, and populations at the two locations were genetically distinct ($\Phi_{ST} = 0.21$). The high diversity at American Samoa indicates that spinner dolphins at this location are not reproductively isolated, but the data do not rule out the possibility that these dolphins may be demographically isolated on ecological timescales.

KEYWORDS: PACIFIC OCEAN; ODONTOCETES; SURVEY – VESSEL; BIOPSY SAMPLING; PHOTO-ID; MOVEMENTS; GENETICS; SPINNER DOLPHINS; ROUGH-TOOTHED DOLPHIN; SPERM WHALE; FALSE KILLER WHALE; BOTTLENOSE DOLPHIN; DWARF SPERM WHALE; SHORT-FINNED PILOT WHALE

INTRODUCTION

The species composition, distribution, abundance and stock structure of odontocete cetaceans in Oceania (the region encompassing the tropical and subtropical waters of the central and western Pacific) are poorly understood, and in many regions, dedicated survey efforts to address these gaps in knowledge have been few. The available information regarding species presence and distribution within this expansive region has recently been compiled and reviewed (see Reeves et al., 1999), and much of the available information has been gleaned from anecdotal reports and opportunistic sightings (e.g. Paton and Gibbs, 2002), historic and recent whaling records (Kasamatsu et al., 1995; Townsend, 1935) and documentation of stranded animals (Trianni and Kessler, 2002). For many portions of this region however, the diversity of cetaceans has never been formally investigated nor have predictions of species presence in certain areas been confirmed.

In particular, very little is known about the odontocetes inhabiting the waters of American Samoa. American Samoa, the only US territory in the Southern Hemisphere, is primarily made up of four eroded volcanic islands (Tutuila, Ofu, Olosega and Ta'u) and two coral atolls/islands (Rose Atoll and Swains Island). Historic whaling records document the catches of sperm whales (*Physeter* macrocephalus) in the waters of American Samoa (Townsend, 1935), and a recent study of humpback whales (Megaptera novaeangliae) in this region indicates their presence during the winter (breeding) period (Robbins and Mattila, 2006). Surveys for cetaceans conducted in nearby Independent Samoa (approximately 95km west of American Samoa) have also confirmed the presence of humpback whales and at least three species of odontocetes thought to inhabit those waters (Noad et al., 2006). However, complementary surveys for odontocetes in the waters of American Samoa are lacking. A recent review of cetacean information for American Samoa (Dolar, 2005) synthesised anecdotal sighting information presented in Reeves et al. (1999) and Craig (2005), species recorded included spotted dolphins (S. attenuata), spinner dolphins (Stenella longirostris), bottlenose dolphins (Tursiops truncatus), rough-toothed dolphins (Steno bredanensis), killer whales (Orcinus orca) and false killer whales (Pseudorca crassidens). In addition, Dolar (2005) noted sightings and strandings of short-finned pilot whales (Globicephala macrorhynchus), a stranded Cuvier's beaked whale (Ziphius cavirostris) and photo-documented spinner dolphins within the coastal waters of Tutuila.

This lack of information on cetaceans in American Samoa has conservation implications. To ensure populations have an acceptable conservation status, resource managers

⁺ Division of Marine Science and Conservation, Nicholas School of the Earth and Environmental Sciences. Duke University Marine Laboratory. 135 Duke Marine Lab Rd., Beaufort, NC 28516, USA.

^{*} Joint Institute for Marine and Atmospheric Research, University of Hawai'i at Manoa. 1000 Pope Rd, Marine Science Building 312, Honolulu, HI 96822, USA.

[#] Provincetown Center for Coastal Studies, 5 Holway Ave., Provincetown, MA 02657, USA.

[~] Hawaiian Islands Humpback Whale National Marine Sanctuary, 726 South Kihei Rd., Kihei, HI 96753, USA.

⁺⁺ Hawai'i Institute of Marine Biology, PO Box 1346, Kane'ohe, HI 96744, USA.

require reliable information on the stock structure and abundance, along with knowledge of any threats they face. Reliable baseline information is also essential to assess and address potential future, and as yet unexpected management issues that may arise. Currently, little information about threats to cetaceans in the waters of American Samoa exists. There is limited coastal development and ecotourism within American Samoa, suggesting that threats from these sources to cetaceans and their habitats are currently minimal. However, there is ample evidence of interactions between cetaceans and commercial fisheries elsewhere in Oceania (see Donoghue et al., 2002) which may represent threats to small, isolated populations if significant bycatch occurs (see Carretta et al., 2005). There is a commercial long-line fishery based in the American Samoa region and only recently has a fisheries observer programme been implemented in this industry. At present, there have been no observed interactions between cetaceans and long-line fishing operations in the waters of American Samoa. Cetaceans are known to interact with longline fisheries operating out of nearby Independent Samoa (Simon Walsh, pers. comm.). An initial review of catch logs for this fishery indicated that approximately 3-6% of sets were affected by depredation, and that false killer whales are associated with damaged catches. Furthermore, rough-toothed dolphins have been seen removing baits from hooks in this fishery.

The purpose of the present study was to conduct the first dedicated surveys for odontocetes in the waters of American Samoa to document the diversity, habitat associations, and stock structure of odontocete cetaceans in this region. For the most abundant species, spinner dolphins, genetic diversity, and allele frequencies of the mitochondrial DNA (mtDNA) control region were compared between American Samoa and the Big Island of Hawaii for a preliminary evaluation of stock structure. Data generated from these surveys will provide a baseline for future efforts to quantify the abundance and structure of management units of cetaceans and assess the sustainability of their populations in the waters of American Samoa.

METHODS

Study site

American Samoa is located approximately halfway between the main Hawaiian Islands and New Zealand in the central Pacific Ocean. The main islands of Tutuila and the Manu'a Islands (Ofu, Olosega and Ta'u) lie at approximately 14°15'S and between 169°24'W and 170°51'W. Maps illustrating the location of American Samoa and its islands are presented in Fig. 1.

Small boat surveys

Between 2003 and 2006, small-boat surveys for cetaceans were conducted in the coastal waters of Tutuila, the main island in American Samoa. Winter/Spring effort (2003-06) was focused on September and early October, coincident with efforts to maximise encounters with humpback whales. Summer surveys (2006) were conducted in February and March. Unsystematic coastal surveys were conducted from 7m to 10m vessels launched from either the north or south side of the island, depending on sea conditions. Two experienced observers, on watch whenever sighting conditions allowed, scanned for cetaceans around the vessel using naked eye and $8 \times$ binoculars. Vessel position was logged by a global positioning system (GPS) at 1-minute or shorter intervals.

Ship surveys

During summer 2006, ship-based surveys were conducted in the coastal waters of the Manu'a Group, Rose Atoll and Swains Island, as well as portions of the offshore regions among these coastal areas. Surveys were conducted from the flying bridge of a 68m oceanographic vessel (NOAA RV Oscar Elton Sette) outfitted with 25×150 'big eye' binoculars. Two observers (port and starboard) scanned constantly from straight ahead to 90 degrees abeam during daylight hours along preset tracklines. Observations were conducted following standard distance sampling/line transect methods for cetaceans, similar to those employed in Barlow (2006). During both small-boat and ship-based surveys, the location, species present, general behavior, and group size estimates for each cetacean encounter were recorded. These sightings were incorporated into a Geographic Information system (GIS) using ArcGIS 9.1 (ESRI, Redlands, CA) for comparison with co-incident depth data. For the small boat survey sightings, gridded (5m cell size) bathymetry of the shelf and slope environments were used (see PIBHMC, 2007). For sightings generated during large ship surveys, a sub-sample of the General Bathymetric Chart of the Oceans (GEBCO) 1-minute worldwide bathymetry dataset was employed to assess habitat associations.

Whenever possible, digital photographs of encountered cetaceans were obtained for species identification confirmations and individual photo-identification. Photographs were obtained with *Canon* digital SLR cameras equipped with 100-300mm zoom lenses. Photographs were shot in 24-bit colour at a resolution of 3072×2048 pixels and saved in jpeg format. All photographs were analysed for quality and were graded based on clarity (1-5), contrast (1-5), and angle of the fin to the photographer (1-5). An overall grade was then assigned to each photograph: 3-8=excellent quality, 9-11=average quality, and \geq 12=poor quality (Friday et al., 2000; Urian et al., 1999). Photographs were then analysed for individual distinctiveness based on the patterns of nicks and notches in the dorsal fins and flukes and, to a lesser extent, by scar or colour patterns on the bodies. Overall distinctiveness was based on a scale of 1 (very distinctive; features evident even in distant or poor quality photograph) to 3 (not distinctive following Friday et al. (2000) and Urian et al. (1999). Those individuals with a distinctiveness rating of 1 or 2 were considered for integration into the photo-identification (ID) catalogue. Photographs of animals were compared between encounters and between years to look for matches. Individuals were entered into the catalogue under one of two conditions; the individual was sighted in more than one encounter and the overall quality of the photograph was at least average, or the individual was sighted once and the overall quality of the photograph was excellent. Where possible, skin/blubber biopsies from animals were obtained from individuals using a Barnett RX-150 crossbow and Ceta-Dart darts with 25mm tips. Biopsy samples were kept on ice and frozen at the end of each day in liquid nitrogen.

Population genetic analyses

Genomic DNA was extracted from 16 tissue samples collected from spinner dolphins in American Samoa using Qiagen *DNEasy* extraction kits. A portion of the mtDNA control region was amplified using the polymerase chain reaction (PCR) and sequenced according to previously described protocols (Andrews *et al.*, 2006). Control region sequences for spinner dolphins sampled at the Big Island of Hawaii (n=18) were obtained from *GenBank* (Accession

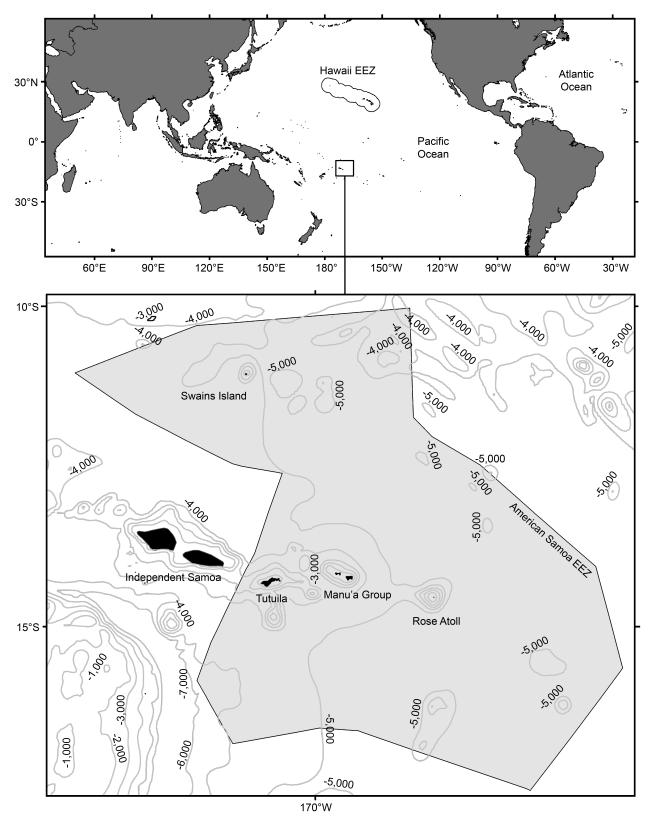


Fig. 1. The location of American Samoa, its Exclusive Economic Zone (EEZ) and its islands in the Central Pacific Ocean.

numbers AY989745-AY989762). All sequences were aligned using SEQUENCHER v.4.2 (Genecodes Corporation), resulting in the comparison of a 390 base pair fragment for all samples.

Nucleotide and haplotype diversities were calculated, and genetic population structure between locations was tested using exact tests of population subdivision (Raymond and Rousset, 1995) and Analysis of Molecular Variance (AMOVA) pairwise Φ_{ST} comparisons. Pairwise Φ_{ST} values can range from 0 to 1, with values of 0 indicating genetic panmixia between locations, and increasing values indicating increasing levels of genetic isolation between locations (Wright, 1951). Significance of pairwise Φ_{ST} comparisons was tested using 100,000 random permutations. All genetic analyses were conducted using ARLEQUIN v.3.11 (Excoffier *et al.*, 2005).

RESULTS

Small boat surveys

During the austral winters of 2003-06, 35 days of surveys were conducted. An average of 87km of trackline per day was covered and a total of 2,900km of trackline was surveyed. Sea state conditions ranged between Beaufort 1 and 4, with most survey effort conducted in Beaufort 2. Thirty groups of cetaceans were encountered during these surveys, producing a daily encounter rate of 1.2 groups of odontocetes per day. During the 2006 austral summer, 11 days of surveys were conducted covering an average of 93km of trackline per day; a total of 1,021.5km of trackline was surveyed. Sea state conditions were optimal in 7 of the 11 surveys days with light winds and Beaufort 0 or 1 conditions throughout the day. The three remaining days had sea states ranging from Beaufort 2 to Beaufort 4. Twentytwo groups of odontocetes were encountered producing a daily encounter rate of 2.2 groups per day. Of the total 3,925km surveyed, approximately 2,200km (56%) were conducted in waters of less than 250m depth.

A total of 52 groups of odontocete cetaceans were encountered during small boat surveys, representing 34 groups of spinner dolphins, 10 groups of rough-toothed dolphins, 1 group of bottlenose dolphins, 2 groups of sperm whales, 3 groups of false killer whales, 1 group of dwarf sperm whales (*Kogia sima*), and 1 group of unidentified odontocetes. Descriptive statistics for odontocete sightings for all small boat surveys are presented in Table 1. Fig. 2 illustrates the tracklines covered during these surveys as well as the locations of cetacean encounters during the austral summer and winter seasons.

Photographs of cetaceans were obtained from each encounter, producing a total of 1,190 frames useful for identifying individual animals within and between survey years. Data on the number of distinct individuals sighted, the number entered into the photo-ID catalogue, and the numbers of resighted individuals are presented in Table 2. The catalogue included 46 spinner dolphins, 41 roughtoothed dolphins, 2 bottlenose dolphins, 5 false killer whales, 1 dwarf sperm whale, and 3 sperm whales. Thirteen (13) spinner dolphins and 14 rough-toothed dolphins were sighted in multiple years. The other catalogued individuals were seen only once or were resighted within the same year. The number of biopsies obtained per species is presented in Table 1.

Ship surveys

During March 2006, six days of surveys were conducted around the Manu'a Islands, Rose Atoll, and Swains Island. Circuit surveys were conducted around each island group. Two circuits were conducted around Rose Atoll and Swains Island; a single circuit and one transect south of the Manu'a Islands were surveyed. Transect lines covered during ship surveys, along with the locations of cetacean sightings are presented in Fig. 3. Approximately 160km of trackline were covered per day, with a total of 960km surveyed. A total of six groups of odontocetes were detected during ship surveys. Two groups of false killer whales and one group of pilot whales were detected in the nearshore waters of the Manu'a Islands; one spinner dolphin was sighted proximate to one of the groups of false killer whales in this region. One group of sperm whales and two groups of unidentified odontocetes were also encountered during ship surveys. Photographs of cetaceans were taken in some of the encounters during the ship surveys, producing 52 frames useful for identifying individuals. As a result, 4 pilot whales and 1 sperm whale were entered into the catalogue.

Population genetic analyses

Nucleotide and haplotype diversities were higher for American Samoa (h = 0.975, $\pi = 0.02$) than for the Big Island of Hawaii (h=0.6993, $\pi = 0.007$). Genetic comparisons between Hawaii and American Samoa were significant for both the exact test of population differentiation (p<0.001) and AMOVA ($\Phi_{ST} = 0.21$, p<0.001).

DISCUSSION

Species diversity

The surveys described here provide baseline data on the diversity of odontocete cetaceans in the waters of American Samoa. Previous reviews of the diversity of cetaceans in the waters of Pacific island nations (Reeves et al., 1999) suggested that at least 15 species were present (some seasonally) in the waters of American Samoa, although at least seven had never been confirmed. Most of these were believed to inhabit the waters of American Samoa based on sightings from other locations in the region with similar habitat envelopes. Several of these species (e.g. bottlenose dolphins and pantropical spotted dolphins) were considered 'common' in American Samoa by Reeves et al. (1999), however, this classification was based on a single reference and may not have accurately represented the most common species found in American Samoa. The results of the surveys described in this paper confirmed the presence of at least seven species of cetaceans in the waters of American Samoa and provided the first confirmed sightings of four species in this region: bottlenose dolphins; false killer whales; rough-toothed dolphins; and dwarf sperm whales.

The results suggest that the cetacean fauna found in American Samoa is indeed similar to that found in the waters of other Pacific island nations. For example,

Table 1
List of odontocete cetaceans encountered during surveys in the coastal waters of Tutuila, American
Samoa including biopsies taken, group size estimates and depth of water.

			Group size estimate			Depth (m)	
Species	Encounters	Biopsies	Mean min.	Mean max.	Range	Mean	SD
Spinner dolphin	34	20	16.7	26.3	2-75	44	21
Rough-toothed dolphin	10	6	13.3	21.1	10-30	123	104
False killer whale	3	5	5	8.3	1-20	229	236
Bottlenose dolphin	1	0	5	10	5-10	90	n/a
Dwarf sperm whale	1	0	4	4	n/a	707	n/a
Sperm whale	2	0	2	2	n/a	907	11
Unidentified odontocete	1	0	4	10	4-10	224	n/a

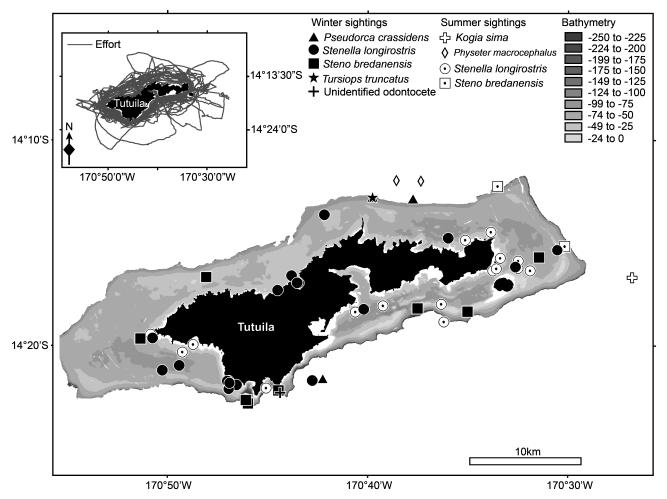


Fig. 2. Survey effort and sighting locations for odontocete cetaceans observed during small boat surveys in the coastal waters of Tutuila, American Samoa during the austral winter (2003-06) and austral summer (2006).

	Table 2		
Distinct individuals determined from Tut	photographs taken du uila, American Samo	Ç ,	e coastal waters of
Total no. distinct	No. entered into	No. resighted	No. resighted withi

Species	Total no. distinct individuals	No. entered into catalogue	No. resighted between years	No. resighted within 2006
Spinner dolphin	131	46	13	8
Rough-toothed dolphin	81	41	14	4
False killer whale	7	5	0	0
Bottlenose dolphin	2	2	0	0
Dwarf sperm whale	3	1	0	0
Sperm whale	3	3	0	0

Noad et al. (2006) found spinner dolphins, false killer whales, and sperm whales during 24 days of joint visual/acoustic surveys in the waters of nearby Independent Samoa (see Fig. 1). Recent joint visual/acoustic surveys in the Solomon Islands (Kahn, 2006) found direct evidence of twelve species of cetaceans. Multiple groups of spinner dolphins, pantropical spotted dolphins, and common bottlenose dolphins were sighted during their surveys, as were single encounters with Indo-Pacific bottlenose dolphins (T. aduncus), killer whales, Risso's dolphins (Grampus griseus), rough-toothed dolphins and shortfinned pilot whales. These surveys also generated single sightings of Mesoplodon beaked whales (Mesoplodon sp.). Sperm whales were detected acoustically on these surveys but were not seen. Unrelated to their directed survey effort, Kahn (2006) also reported on a single stranded false killer

whale whose skeleton was found on a remote beach within the Arnavon Islands Marine Protected Area. False killer whales had been previously sighted near the Solomon Islands (Shimada and Pastene, 1995).

Habitat use

During the small-boat surveys around Tutuila, three out of the six confirmed species (false killer whales, dwarf sperm whales and sperm whales) were encountered in waters deeper, on average, than 200m (Table 1). Both sperm whale species are deep diving odontocetes and are commonly found in abyssal waters (Baird, 2005; Watkins *et al.*, 2002). While false killer whales are not known to be deep divers (e.g. Ligon and Baird, 2001) they are frequently sighted in waters deeper than 200m in other portions of their range. Ship-based surveys around the Manu'a Islands, Rose Atoll

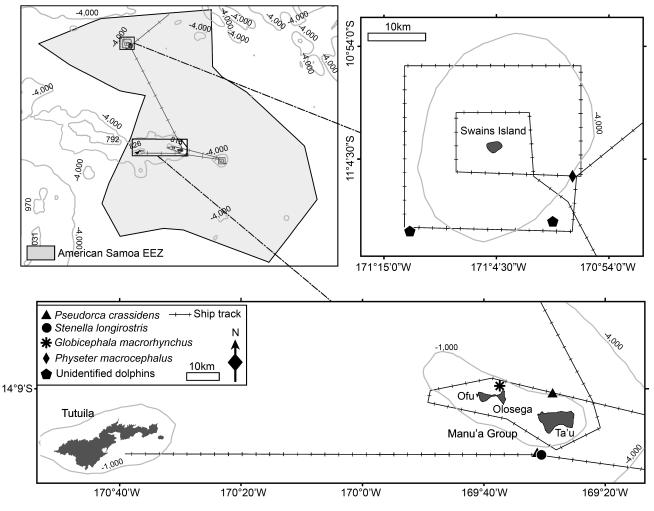


Fig. 3. Survey tracklines and sighting locations for odontocete cetaceans observed during ship-based surveys in the waters of American Samoa during the austral summer 2006.

and Swains Island were biased toward deeper water habitats. For example, sperm whales were encountered in 4,055m deep water, false killer whales were encountered in $1,040\pm374m$ deep water and pilot whales were encountered in 375m deep water.

Spinner dolphin encounters were generally inshore (often in or near bays and over regions of sandy bottom less than 60m deep), despite substantial survey effort in waters deeper than 120m. These daytime, inshore sighting patterns are indicative of behaviors exhibited by island-associated spinner dolphins elsewhere in the Pacific. For example, in Hawaii (Karczmarski et al., 2005; Norris et al., 1994), French Polynesia (Poole, 1995) and the Solomon Islands (Kahn, 2006), spinner dolphins use sheltered inshore regions during the day for resting and predator avoidance. The inshore daytime habits of these animals often make them the focus of tourism operations (Norris et al., 1994) and there is growing concern in some areas that intensive tourism may present a threat by impinging on their resting behavior or excluding them from primary resting habitats. There are currently no dedicated dolphin-watching operations in American Samoa.

Stock structure

For management purposes, cetaceans inhabiting the waters of American Samoa are considered *protected species* under the US Marine Mammal Protection Act (MMPA). The MMPA requires that assessments for each stock of marine mammals be conducted. These assessments include details on population discreteness and abundance estimates. In several locations in the Pacific Islands, cetaceans often exhibit island-associated population discreteness, with restricted gene flow at meso-spatial scales. For example, spinner dolphins exhibit restricted gene flow between islands separated by as little as 17km in the Society Islands (Oremus *et al.*, 2007) and as little as 50km in the Hawaiian Archipelago (Andrews *et al.*, 2006). Bottlenose dolphins also exhibit restricted gene flow between islands in the Hawaiian Archipelago (Martien *et al.*, In review). On a larger geographic scale, island-associated false killer whales in Hawaii appear genetically distinct from those found in the Eastern Tropical Pacific and around Palmyra Atoll (Chivers *et al.*, 2007).

Consistent with observations at other locations in the Pacific, the photographic identification data provide preliminary evidence for population discreteness for some cetacean species at the islands of American Samoa. The relatively high resighting rates of both rough-toothed and spinner dolphins in the waters of Tutuila, and the spatial proximity of between-year sightings for some individuals (e.g. one individual rough-toothed dolphin sighted in 2004 was resighted in 2006 approximately 2km away from the initial sighting), suggest some level of site fidelity for these two species. However, if cetacean populations were completely isolated at American Samoa, small population sizes and low genetic diversity could be predicted based on

the small size of this geographic region. Contrary to this prediction, spinner dolphin genetic diversity at American Samoa is higher compared to the Big Island of Hawaii, despite the fact that the Big Island has more available resting habitat than American Samoa. Therefore, the high genetic diversity is likely not the result of a larger population size at American Samoa but rather the result of gene flow between American Samoa and other locations. However, the level of gene flow may be low enough that spinner dolphins at American Samoa remain demographically isolated on ecological time scales that are relevant to conservation. For example, in the Society Archipelago, spinner dolphins at different islands have high genetic diversity despite demographic closure, as revealed by photo-ID data (Oremus et al., 2007). The high diversity is hypothesised to be due to metapopulation dynamics, with occasional inter-island movement by 'visitors' and long-term migrants between islands across a geographic area larger than the Society Archipelago. The genetic and photo-ID data presented, while preliminary, follow a pattern similar to that in the Society Archipelago, indicating that metapopulation dynamics may be occurring similarly in American Samoa.

The genetic population structure analyses provide further support for the hypothesis of metapopulation dynamics for spinner dolphins at American Samoa. The exact test and pairwise Φ_{ST} test comparing American Samoa and the Big Island of Hawaii ($\Phi_{ST} = 0.21$) indicate that these locations are genetically distinct. This result is not surprising given that genetic distinctions were found between spinner dolphins at different islands within Hawaii separated by as little as 50km (Andrews et al., 2006). However, the pairwise $\Phi_{\rm ST}$ value between American Samoa and the Big Island is similar to Φ_{ST} values for the same genetic locus between spinner dolphins at Pacific islands separated by much less geographic distance. For example, pairwise Φ_{ST} values within the Hawaiian Archipelago were as high as 0.16 (K. Andrews, unpublished data), and pairwise Φ_{ST} values between islands within the Society Archipelago were as high as 0.32 (Oremus et al., 2007). The fact that American Samoa and the Big Island of Hawaii have a comparable pairwise Φ_{ST} despite the large geographic distance separating these islands indicates that some gene flow may occasionally occur between islands within a larger region of the Pacific Islands, including these two locations. However, consistent with the predictions of metapopulation dynamics, this gene flow appears to be low enough that spinner dolphins at these islands are demographically discrete on an ecological timescale and could be considered separate management units for conservation. Collection of more genetic and photographic identification data is needed at American Samoa and other locations in the Pacific to determine the extent of genetic and demographic connectivity between American Samoa and other locations. Similar data are needed for each cetacean species occurring at American Samoa before effective management decisions can be made concerning stock structure.

CONCLUSIONS

These surveys provide data on the cetacean species diversity in American Samoa, information that is critical for marine mammal management purposes in the US. Further surveys are required to fully assess the diversity of cetaceans and to develop abundance estimates for odontocetes in American Samoa waters. Further genetic and photo-ID studies will help establish the relationships between cetaceans sampled in this region and those found in the waters of other nearby Pacific Island Region nations. In particular, coordinated surveys with researchers in Independent Samoa would be extremely useful for stock assessments of several of the island-associated species such as spinner dolphins and bottlenose dolphins. Similarly, further research is required to assess the potential threats to cetaceans in waters of American Samoa.

ACKNOWLEDGMENTS

Thanks to Jason Baker, Chad Yoshinga, Karolyn Braun, Siri Hakala, Lisa Conger, Elliott Hazen, Hugh Finn, Tamara Maguire, Shannon Rankin, Amy Kennedy, Ed Lyman and Juney Ward during field efforts in American Samoa. Thanks to R.L. Pitman and L. Galver for collecting and analyzing the GenBank samples used in this study. As well, we would like to thank Ruth Utzurrum (American Samoa Division of Marine and Wildlife Resources), Nancy Daschbach (Fagatele Bay National Marine Sanctuary) and Peter Craig (US National Park Service) for logistical help in American Samoa. Earlier drafts of this manuscript were improved by comments by Jason Baker, Charles Littnan and Kerry Irish. This research was conducted under US MMPA/ESA permit 774-1714-03 and Scientific Collection Permits from the American Samoa Department of Marine and Wildlife Resources. All research conducted conformed to University of Hawaii IACUC regulations.

REFERENCES

- Andrews, K.R., Karczmarski, L., Au, W.L., Rickards, S.H., Vanderlip, C.A. and Toonen, R.J. 2006. Patterns of genetic diversity of the Hawaiian spinner dolphin (*Stenella longirostris*). *Atoll Res. Bull.* 543: 65-73.
- Baird, R. 2005. Sightings of dwarf (*Kogia sima*) and pygmy (*K. breviceps*) sperm whales from the main Hawaiian Islands. *Pac. Sci.* 59: 461-66.
- Barlow, J. 2006. Cetacean abundance in Hawaiian waters estimated from a summer/fall survey in 2002. Mar. Mammal Sci. 22(2): 446-64.
- Carretta, J., Forney, K., Muto, M., Barlow, J., Baker, J., Hanson, B. and Lowry, M. 2005. US Pacific Marine Mammal Stock Assessments: 2004. NOAA Technical Memorandum NMFS SWFSC-375(May 2005). 315pp.
- Chivers, S., Baird, R.W., McSweeney, D.J., Webster, D.L., Hedrick, N.M. and Salinas, J.C. 2007. Genetic variation and evidence for population structure in eastern North Pacific false killer whales (*Pseudorca crassidens*). Can. J. Zool. 85: 783-94.
- Craig, P. 2005. Natural History Guide to American Samoa. National Park of American Samoa, Pago Pago, American Samoa. [Available from www.nps.gov].
- Dolar, M.L.M. 2005. Cetaceans of American Samoa. Report submitted to the Department of Marine and Wildlife Resources, American Samoan Government, June 2005 (unpublished). 24pp.
- Donoghue, M., Reeves, R. and Stone, G. 2002. Report of the Workshop on Interactions Between Cetaceans and Longline Fisheries. Apia, Samoa, November 2002, New England Aquarium Aquatic Forum Series Report. 03-1. New England Aquarium Press, Boston, USA. 45pp.
- Excoffier, L., Laval, G. and Schneider, S. 2005. Arlequin ver. 3.0: an integrated software for popultion genetics data analysis. *Evol. Bioinf. Online* 1: 47-50.
- Friday, N., Smith, T., Stevick, P. and Allen, J. 2000. Measurement of photographic quality and whale distinctiveness for the photographic identification of humpback whales. *Mar. Mammal Sci.* 16(2): 355-74.
- Kahn, B. 2006. Oceanic cetaceans and associated habitats. In: Green, A., Lokani, P., Atu, W., Ramohia, P., Thomas, P. and Almany, J. (eds). Solomon Islands Marine Assessment: Technical Report of survey conducted May 13 to June 17, 2004. TNC Pacific Island Countries Report No. 1/06. The Nature Conservancy, South Brisbane, QLD.
- Karczmarski, L., Wursig, B., Gailey, G., Larson, K.W. and Vanderlip, C.A. 2005. Spinner dolphins in a remote Hawaiian atoll: social grouping and population structure. *Behav. Ecol.* 16: 675-85.
- Kasamatsu, F., Nishiwaki, S. and Ishikawa, H. 1995. Breeding areas and southbound migrations of southern minke whales *Balaenoptera* acutorostrata. Mar. Ecol. Prog. Ser. 119(1-3): 1-10.

- Ligon, A. and Baird, R. 2001. Diving behaviour of false killer whales off Maui and Lana'i, Hawaii. Presented to the 14th Biennial Conference on the Biology of Marine Mammals, Vancouver, Canada, December 2001.
- Martien, K.K., Baird, R.W., Hedrick, N., Gorgone, A.M., Lowther, J., McSweeney, D., Robertson, K. and Webster, D.L. In review. Population structure of bottlenose dolphins around the main Hawaiian Islands revealed by mitochondrial and microsatellite markers. *Can. J. Zool.*
- Noad, M.J., Paton, D.A., Gibbs, N.J. and Childerhouse, S.J. 2006. A combined visual and acoustic survey of humpback whales and other cetaceans of Samoa. Paper SC/A06/HW28 presented to the IWC Workshop on Comprehensive Assessment of Southern Hemisphere Humpback Whales, Hobart, Tasmania, 3-7 April 2006 (unpublished). 15pp. [Paper available from the office of this Journal].
- Norris, K.S., Würsig, B., Wells, R.S. and Würsig, M. 1994. *The Hawaiian Spinner Dolphin*. University of California Press, Berkeley, California. 408pp.
- Oremus, M., Poole, M.M., Steel, D. and Baker, C.S. 2007. Isolation and interchange among insular spinner dolphin communities in the South Pacific revealed by individual identification and genetic diversity. *Mar. Ecol. Prog. Ser.* 336: 275-89.
- Paton, D. and Gibbs, N. 2002. Anecdotal and reported sightings of cetaceans with the Samoan, Fiji, Vanuatu and Solomon Island Regions. Report to Environment Australia. Australian Government Department of Environment and Heritage, Canberra, ACT.
- PIBHMC. 2007. Gridded bathymetry of Tutuila Island, American Samoa. Pacific Islands Benthic Habitat Mapping Center Coral Reef Ecosystem Division, PIFSC, NOAA and the Joint Institute for Marine and Atmospheric Research (JIMAR). [Available at: http://soest.hawaii.edu/pibhmc].
- Poole, M.M. 1995. Aspects of behavioural ecology of spinner dolphins (*Stenella longirostris*) in the nearshore waters of Mo'orea, French Polynesia. PhD dissertation, University of California at Santa Cruz. 177pp.

- Raymond, M. and Rousset, F. 1995. An exact test for population differentiation. *Evolution* 49: 1280-83.
- Reeves, R., Leatherwood, S., Stone, G.S. and Eldredge, L.G. 1999. Marine mammals in the area served by the South Pacific Regional Environment Programme (SPREP). South Pacific Regional Environment Programme, Apia, Samoa.
- Robbins, J. and Mattila, D.K. 2006. Summary of humpback whale research at American Samoa, 2003-2005. Paper SC/58/SH5 presented to the IWC Scientific Committee, May 2006, St. Kitts and Nevis, West Indies (unpublished). 4pp. [Paper available from the Office of this Journal].
- Shimada, H. and Pastene, L.A. 1995. Report of a sighting survey off the Solomon Islands with comments on Bryde's whale distribution. *Rep. int. Whal. Commn* 45: 413-18.
- Townsend, C.H. 1935. The distribution of certain whales as shown by logbook records of American whaleships. *Zoologica (NY)* 19(1-2): 1-50+6 maps.
- Trianni, M.S. and Kessler, C.C. 2002. Incidence and strandings of the spinner dolphin, *Stenella longirostris*, in Saipan Lagoon. *Micronesia* 34: 249-60.
- Urian, K., Hohn, A.A. and Hansen, L.J. 1999. Status of the photoidentification catalog of coastal bottlenose dolphins of the western North Atlantic. Report of a workshop of catalog contributors. NOAA Technical Memorandum NMFS-SEFSC 425. 22pp. [Available from: www.nmfs.gov].
- Watkins, W.A., Daher, M.A., DiMarzio, N.A., Samuels, A., Wartzok, D., Fristrup, K.M., Howey, P.W. and Maiefski, R.R. 2002. Sperm whale dives tracked by radio tag telemetry. *Mar. Mammal Sci.* 18(1): 55-68.
- Wright, S. 1951. The genetic structure of populations. Ann. Eugenics 15: 323-54. IIII.

Received: November 2007 Accepted: December 2007